

SHAWURENSINE, A NEW C₁₉-DITERPENOID ALKALOID FROM *Delphinium shawurense*

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*The new norditerpenoid alkaloid shawurensine in addition to β -sitosterol and the known alkaloids elatine, delbrulin, and methyllycaconitine were isolated from *Delphinium shawurense*. A structure for shawurensine was proposed based on spectral data.*

Key words: *Delphinium shawurense*, Ranunculaceae, C₁₉-norditerpenoid alkaloid, shawurensine.

Plants of the genus *Delphinium* (Ranunculaceae) are sources of diterpenoid alkaloids. Therefore, many species of this genus are used in folk medicine [1]. However, data on the chemical composition and use of *D. shawurense* W. T. Wang in folk medicine have not been published. The plant was collected in Xinjiang Autonomous Republic of the Chinese People's Republic during flowering in July 2005.

We investigated the alkaloid composition of the aerial part of *D. shawurense*. Ethanol extraction produced the total alkaloids, the yield of which was 0.56% of the air-dried plant mass. The known C₁₉-norditerpenoid alkaloids elatine, delbruline, and methyllycaconitine and the new base **1**, which we called shawurensine, in addition to β -sitosterol were isolated from the total alkaloids. The known compounds were identified by comparing their spectral properties with those published [2-5].

Shawurensine (**1**) was isolated as an amorphous white powder with molecular formula C₃₇H₃₂N₂O₁₁ as determined by high-resolution mass spectrometry (HR-ESI) and confirmed by PMR and ¹³C NMR spectra.

The PMR spectrum of **1** contained signals for protons of *N*-ethyl, four methoxyls, 18- and 19-methylenes, NH groups, and *o*-substituted benzene ring (Table 1). The presence of two N atoms in **1** and the spectral data given above enable **1** to be classified as a C₁₉-norditerpenoid alkaloid with a C-18 ester.

Chemical shifts (CS) and multiplicities of proton signals at 3.92 ppm (br) and 3.61 (3H, t, J = 4.8 Hz) in the PMR spectrum of **1** confirmed the presence of C-6 and C-14 methoxyls, respectively [6, 7]. The ¹³C NMR spectrum contained signals for 37 C atoms and confirmed the conclusions made on the basis of the mass and PMR spectra of **1** (Table 2). Furthermore, the ¹³C NMR spectrum had signals at 90.8, 88.3, and 77.4 ppm, confirming the presence of a C-6 methoxyl and C-7 and C-8 hydroxyls [7]. CS in the ¹³C NMR spectrum of **1** at 83.9 and 55.8 ppm in addition to 82.5 and 56.3 ppm were consistent with C-1 and C-16 methoxyls.

The molecular formula of **1** includes 11 O atoms from 4 methoxyls, an ester, 2 hydroxyls, and 2 carbonyls. Therefore, the last O atom belongs to a carboxyl group, which was confirmed by comparing the ¹³C NMR spectra of **1** and delavaine A (**2**) [8], which showed that the CS of the C atoms were similar and differed only by an additional signal for OCH₃ at 51.7 ppm in the spectrum of **2**. The compositions differed only by a CH₂ group. The PMR spectrum of **1** had signals at 3.12 ppm (1H, br.s, CH-CH₃), 2.96 (2H, br.s, CH₂COOH), and 1.31 (3H, d, J = 7.2 Hz, CH-CH₃), which were due to a [*N*-(2''-methylsuccinyl)anthranyl] moiety in **1**. Analogous proton signals were observed in spectra of elatine [2], geyerline [9], and grandiflorine [9], which were due to proton signals of the [*N*-(2''-methylsuccinimido)anthranyl] moiety in these alkaloids. The PMR spectrum of **1** differed from those of elatine, geyerline, and grandiflorine by an NH signal, which was consistent with the presence in **1** of an [*N*-(2''-methylsuccinyl)anthranyl] moiety. Therefore, **1** had the structure 1.

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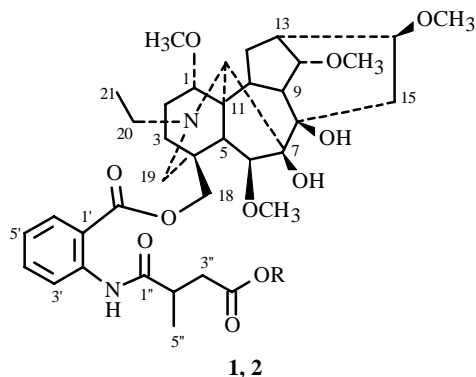
TABLE 1. Chemical Shifts in PMR Spectrum of Shawurensine (600 MHz, CDCl₃, 0 = TMS, δ, ppm, J/Hz)

Atom	1	Atom	1
1	3.00 (1H, m)	17	2.63 (1H, br.s)
2a	2.09 (1H, m)	2H-18	4.18 (2H, br.s)
2b	2.17 (1H, m)	19a	2.48 (1H, d, J = 11.4)
3a	1.53 (1H, m)	19b	2.73 (1H, d, J = 11.4)
3b	1.80 (1H, d, J = 15)	Ar-H-3'	7.95 (1H, d, J = 8.4)
5	1.73 (1H, br.s)	H-4'	7.09 (1H, t, J = 7.7)
6a	3.92 (1H, br.s)	H-5'	7.52 (1H, t, J = 7.5)
9	2.97 (1H, d, J = 7.2)	H-6'	8.68 (1H, d, J = 8.4)
10	1.95 (1H, q, J = 12.0, J = 6.0)	CO-NH	11.09 (1H, br.s)
12a	1.67 (1H, dd, J = 15.0 and 6.6)	2''	3.12 (1H, br.s)
12b	2.43 (1H, dd, J = 15.0 and 8.4)	3''	2.96 (2H, br.s)
13	2.35 (1H, dd, J = 6.6 and 4.8)	5''-CH ₃	1.31 (3H, d, J = 7.2, N-CH ₂ CH ₃)
14b	3.61 (1H, t, J = 4.5)	N-CH ₂ CH ₃	2.84-2.81 (2H, m)
15a	1.85 (1H, dd, J = 14.4 and 7.8)	N-CH ₂ CH ₃	(3H, t, J = 7.2)
15b	2.43 (1H, dd, J = 15.0 and 8.4)	4×OCH ₃	3.40, 3.37, 3.34, 3.26 (3H each, s)
16a	3.08 (1H, t, J = 6.0)		

TABLE 2. Chemical Shifts in ¹³C NMR Spectrum of Shawurensine (1) and Delavaine A (2) (600 MHz, CDCl₃, 0 = TMS, δ, ppm)

C atom	1	2	C atom	1	2
1	83.9	83.8	20	50.9	50.9
2	25.9	25.9	21	14.0	13.9
3	32.0	31.9	1'	114.9	114.6
4	37.5	37.5	2'	141.6	141.4
5	43.3	43.1	3'	120.7	120.5
6	90.8	90.8	4'	134.8	134.9
7	88.5	88.3	5'	122.6	122.5
8	77.5	77.4	6'	130.3	130.2
9	50.4	50.2	1''	174.8	172.4
10	38.4	38.0	2''	39.1	38.9
11	49.0	79.0	3''	37.0	37.5
12	28.6	28.6	4''	176.0	174.0
13	45.9	46.0	5''	17.9	17.9
14	83.9	83.8	1-OCH ₃	55.8	55.8
15	33.6	33.5	6-OCH ₃	57.8	57.8
16	82.5	82.5	14-OCH ₃	58.1	58.0
17	64.5	64.4	16-OCH ₃	56.3	56.2
18	69.8	69.7	ArC=O	167.9	167.9
19	52.3	52.4	COOCH ₃	-	57.1

The mass spectrum of **1** had a peak for the molecular ion with m/z 701 [$M + 1$]⁺ and a peak at m/z 587 that was probably formed by cleavage of a water molecule from the HN-CO-CH(CH₃)-CH₂-COOH moiety to form C₅H₆NO₂ whereas the peak with m/z 651 was apparently formed by simultaneous elimination of OCH₃ and H₂O from the molecular ion.



1: R = H; 2: R = CH₃

EXPERIMENTAL

General Comments. Melting points were determined on a MP-J3 apparatus (Yanaco). Mass spectra and elemental analyses were obtained in a FT ICR MS IonSpec 7.0T (USA) spectrometer. PMR and ¹³C NMR spectra were recorded on Bruker Avance 600 NMR spectrometers relative to TMS (internal standard). ¹³C NMR spectra were obtained with full C–H decoupling. TLC was performed on HSGF254 silica-gel plates (Qingdao Hai Yang Chemical Co., Ltd., Qingdao, China). Compounds on silica-gel plates were developed by iodine vapor, a fluorescent lamp, and spraying with Dragendorff's reagent. Column chromatography used silica gel (200-300 mesh, Qingdao, China).

Plant Material. The aerial part of *D. shawurense* W. T. Wang was identified by Prof. Shi Ming Duan of Xinjiang Institute of Ecology and Geography.

Extraction and Isolation of Alkaloids. Dried and ground raw material (2.9 kg) was extracted exhaustively with ethanol (80%) at room temperature. The extract was condensed in vacuo. The aqueous residue was diluted with H₂SO₄ solution (5%). The acidic solution was filtered and washed with CHCl₃.

The CHCl₃ extract was washed with saturated carbonate solution and water. The solvent was removed in vacuo to afford a mixture of alkaloids (6.3 g).

The acidic aqueous solution was made basic with carbonate solution until the pH was 8. Alkaloids were extracted with CHCl₃. Solvent was distilled off in vacuo to afford a mixture of alkaloids (10.1 g). The overall yield was 16.4 g or 0.56% of the dry starting material.

The alkaloid fraction (6.3 g) was worked up successively with boiling petroleum ether and acetone to afford fractions that were soluble and insoluble in acetone. The acetone-soluble fraction afforded elatine (1.0 g). The mother liquor from elatine was chromatographed over a column of silica gel with elution by petroleum ether:acetone (7:3 and 1:1). Preparative TLC of fractions 8-31 on silica-gel plates using hexane:CHCl₃:CH₃OH (7:3:1) produced methyllycaconitine (40 mg).

The acetone-insoluble part was fractionated over a column of LH-20 Sephadex with elution by CH₃OH. Fractions 4-9 were chromatographed over a column of silica gel with elution by petroleum ether:CHCl₃:CH₃OH (30:40:4 and 30:18:6) to afford amorphous **1** (20 mg).

The other alkaloid fraction (10.1 g) was separated by column chromatography over silica gel with gradient elution by petroleum ether:acetone (10:1→1:1). The first five fractions afforded β-sitosterol; fractions 33-37, delbruline (120 mg).

Shawurensine (1), C₃₇H₅₂N₂O₁₁, amorphous base. Mass spectrum (HRESI, *m/z*, %): 701 (100) [M + 1]⁺, 651 (25), 587 (56.2), 273 (9.3), 219 (9.3).

Tables 1 and 2 give the PMR and ¹³C NMR spectra of **1**.

Elatine (2), C₃₈H₅₀N₂O₁₀, mp 225-227°C (acetone). Mass spectrum (FAB—MS): 695 [M + H]⁺.

PMR spectrum (400 MHz, CDCl₃, δ, ppm, J/Hz): 8.05 (1H, d, J = 7.6, Ar–H-6'), 7.67 (1H, dt, J = 7.6, 1.6, Ar–H-5'), 7.53 (1H, dt, J = 7.6, 1.6, Ar–H-4'), 7.27 (1H, d, J = 7.6, Ar–H-3'), 5.07 (2H, s, OCH₂O), 4.12-4.02 (2H, m, H-18), 3.70 (1H, t, J = 5.6, H-14β), 3.64 (1H, s, H-6α), 3.26, 3.32, 3.34, 3.43 (3H each, s, 4 × OCH₃), 2.86-2.79 (1H, m, NCH₂b), 2.76 (1H, d, J = 11.6, H-19a), 2.73-2.63 (1H, m, NCH₂a), 2.58 (1H, dd, J = 14.0, 4.4, H-12a), 2.44 (1H, dd, J = 14.8, 8.4, H-15a), 2.40-2.30

(2H, m, H-19b, H-13), 1.87 (1H, dd, J = 14.8, 8.0, H-15b), 1.78-1.69 (1H, m, H-12b), 1.07 (3H, t, J = 7.2, NCH₂CH₃), 3.05 (1H, s, Suc-CH), 2.14 (m, Suc-CH₂), 1.47 (3H, d, J = 7.2, Suc-CH₃).

These spectral properties are identical to those published for elatine [2, 7].

Delbruline, C₂₆H₄₁NO₇, mp 142-143°C (acetone). Mass spectrum (ESI, *m/z*, %): 480.0 (100) [M + 1]⁺, 292.0 (5.6), 240.1 (6.3), 229.1 (13.8), 207.2 (50).

The PMR and ¹³C NMR spectra were identical to those published for delbruline [3].

Methyllycaonitine, C₃₇H₅₀N₂O₁₀, amorphous compound. Mass spectrum (ESI, *m/z*, %): 683.2 (88.2) [M + 1]⁺, 665.2 (20.5) [M - 17]⁺, 651.1 (100) [M - 31]⁺, 633.1 (23.5) [M - 49]⁺, 619.2 (69.4), 601.2 (23.5), 573.2 (17.6), 386.1 (11.8), 354.2 (10).

PMR spectrum (600 MHz, CD₃COCD₃, δ, ppm, J/Hz): 8.26 (1H, d, J = 7.8, Ar-H-6'), 7.86 (1H, t, J = 7.8, Ar-H-5'), 7.73 (1H, t, J = 7.8, Ar-H-4'), 7.49 (1H, d, J = 7.8, Ar-H-3'), 4.31-4.20 (2H, m, H-18), 4.05 (1H, s, H-6α), 3.69 (1H, t, J = 4.8, H-14β), 3.52, 3.41, 3.37, 3.36 (3H, s, 4 × OMe), 3.25 (1H, t, J = 7.8, H-16α), 2.86 (1H, d, J = 11.4, H-19a), 2.72-2.67 (2H, m, NCH₂CH₃), 1.15 (3H, t, J = 7.0, NCH₂CH₃), 3.05 (1H, s, Suc-CH), 2.38 (2H, m, Suc-CH₂), 1.50 (3H, d, J = 7.2, Suc-CH₃).

The spectral properties were identical to those published for methyllycaonitine [4, 7].

β-Sitosterol, mp 135-136°C (acetone).

The spectral properties were identical to those published for β-sitosterol [5].

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